Isolation and evaluation of bacterial biocontrol agents for management of leaf spot (Myrothecium roridum) in mulberry

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Biocontrol agents (BCAs) are promising alternatives to chemicals in crop protection. Watery extracts of various composts and associated bacteria are found success to control various diseases of agricultural crops. Studies were conducted to identify potential BCAs from various compost extracts to control leaf spot caused by *Myrothecium roridum* in mulberry (*Morus* spp.). Twelve numbers of bacteria isolated from watery extracts of various composted materials were tested for their ability to suppress the disease and compared with the efficacy of carbendazim (0.1%), a recommended fungicide for management of the disease. *In vitro* studies showed significant (P < 0.05) suppression of mycelia growth and sporulation of the pathogen by six bacterial isolates in potato dextrose broth. To test their *in vivo* efficacy, experiments were conducted on mulberry plants (cv. S_1) raised in earthen pots spraying with 48 h old bacterial suspension (1×10^8 CFU/ml) at 30 days after pruning. Disease severity recorded 7, 14 and 21 day after treatment revealed significant (P < 0.01) influence of three *Bacillus* isolates *viz. B. lentimorbus, B. pumilus* and *B. cerceus* reducing the disease severity comparable to 0.1% carbendazim. No significant interaction was noticed between bacterial isolates and days after application. Bioassay by feeding silkworms with the leaves 10 days after treatment revealed BCAs are safe to silkworms.

Key words: Biological control agents, leaf spot, mulberry, Myrothecium roridum

INTRODUCTION

Leaf spot caused by Myrothecium roridum (Tode) Ex Fr. is an important disease of mulberry in mulberry growing areas West Bengal. The disease is more frequent during July-October, with maximum severity in August (Prathess Kumar, 2002; Pratheesh Kumar and Chattopadhyay, 2002). The disease causes disruption in the metabolic activities of the host leading to reduction in leaf yield and nutritive value (Umesh Kumar, 1991; Shree and Nataraja, 1993; Pratheesh Kumar et al., 2002). Besides consumption of diseased leaves lead to prolonged larval period, poor cocoon formation, reduced silk content and slik filament length (Qadri et al., 1999). Although the disease can be partially controlled by application of chemical fungicides (Govindaiah et al., 1988; Philip et al., 1990; Teotia et al., 1994), this is not economically viable besides environmental considerations. In addition, repeated use of fungicides may cause development of

resistant pathogen strains. Alternative methods are therefore needed for its management.

There are reports on use of aqueous extracts of compost for foliar disease control (Weltzine, 1991; Trankner, 1992; Jongebloed et al., 1993; Elad and Steinberg, 1994). Weltzien observed good control of gray mould (Botrytis cinerea) on strawberries, various powdery mildews, and late blight of potato (Phytophthota infestans) upon treating composted plant extracts. The mechanism of phytosanitary effect of decomposed plant extracts is attributed to biological activity of microorganisms available in the extracts. Similar experiments resulted control of powdery mildew in mulberry by watery extracts of various composts (Pratheesh Kumar et al., 2000). Present experiments were conducted to assess the bacteria isolated from various composted organic materials, which are found effective for the control of leaf spot caused by Myrothecium roridum in mulberry.

MATERIALS & METHODS

Twelve bacterial strains were isolated from compost of silkwrom rearing waste, Eichhornia, and compost of common weeds, which are used as organic manure in mulberry cultivation. The potato dextrose broth was prepared and poured in 250 ml conical flask. All the twelve bacterial isolates after 24 h growth in nutrient medium (peptone, 5 g; beef extract, 3 g; NaCl, 5 g; agar, 5 g distilled water, 11) were inoculated separately in the conical flask containing broth. A 5 mm viable disc of mycelia obtained from 7 day old culture of test fungi was transferred in each conical flask. Broth with carbendazim 0.1% was kept for comparison. Similarly, the broth without bacterial isolate was also kept as check. The treatments and control was replicated thrice. The whole set was incubated at 28 ± 2°C. Fourteen days after incubation the spores were harvested and enumerated using hemocytometer. Similarly, the mycelia were harvested dried at 60°C for 72 h and weighed.

Two sets of experiments were conducted for in vivo evaluation of antagonistic bacterial isolates. The bacterial isolates were grown on soild nutrient medium after 24 h of growth; the bacterial suspension was prepared in sterilized distilled water and adjusted to an absorbance of 0.1 at 600 nm (108 CFU/ml). These suspensions were then sprayed separately on 30 day old mulberry plants (cv. S₁) raised in earthen pots following the recommended package of practices (Subba Rao, 1989) with a hand held sprayer. Recommended fungicides carbendazim (0.1%) was sprayed separately. A control row, which was sprayed with sterilized distilled water, was maintained for comparison. Conidial suspension (1× 106 / ml) was prepared in distilled water using one-week-old culture of Myrothecium roridum grown in potato dextrose agar. The suspension was sprayed on the mulberry plants 24 h after treatment. Three replications were kept for each treatment and control in random. The data on the severity of leaf spot was recorded in a 0-5 scale where, 0 = no infection, 1 = 1-5%, 2 = 6-15 %, 3 = 16-30 %, 4 = 31-50 % and 5 = > 50 % leaf area infected with leaf spot. Severity (%) was then calculated using formula

Disease severity (%) =
$$\frac{\Sigma \text{ Numerical values}}{\text{Total number of leaves}} \times 100$$
observed × maximum score (5)

A third set of experiment was conducted by using only four identified effective bacterial isolates *viz*. Arthrobacter ilicis, Bacillus cereus. B. lentimorbus and B. pumilus., identified from the previous experiment. Experimental procedures were followed as in the previous experiments except disease severity was recorded 7, 14 and 21 days after treatment.

Bioassay was conducted following the method (Jolly, 1987). Multivoltine silkworms (Nistari × Nistari) were fed with leaves harvested 10 days after spray of the bacterial strains of 30 days old mulberry plants (cv. S1). Single cocoon weight, single shell weight, effective rearing rate, 10 mature larval weight, silk ratio and total silk filament length were calculated following standard methods.

The data were subjected for ANOVA, F test were conducted and the values were compared for least significant difference.

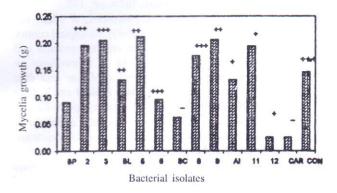
RESULTS AND DISCUSSION

Bacterial isolates significantly influenced on the mycelia production. Six bacterial isolates and carbendazim showed significant reduction in fungal growth. Interestingly, except two bacterial isolates, spore production was significantly decreased in all other treatments (Fig. 1). In the first set of in vivo experiment with all the bacterial isolates, four isolates significantly reduced the disease severity > 40 % compared with control. However, carbendazim more effectively controlled the disease. The effective four bacterial strains were identified as Arthobacter ilicis, Bacillus lentomorbus, B. cercus, and B. pumilus which reduced the disease severity from 14.7 % to 8.2 %, 6.8, 7.8 % (Fig. 2), respectively controlling 44.1 %, 53.8 %, 50.7 % and 47.2 % fourteen days after treatment.

In the second set of experiment, all bacterial isolates reduced disease severity. In all the treatment maximum reduction in disease severity was obtained 21 days after treatment. Here also

carbendazim reduced the severity maximum (66.7 %) followed by *B. lentimorbus* (53.5 %), *B. pumilus* (48.6 %), *B. cereus* (46.1 %) and *A. ilicis* (31.6 %).

Myrothecium roridum



Mean of three replications, BP—B. pumilus, BL—B. lentimorbus, BC—B. cereus, AI—A. ilicis, CAR—carbendazim, CON—control. +++ high, ++ medium, + low and - no sporulation.

Fig. 1: Effect of bacterial isolates on mycelia growth and sporulation of Myrothecium roridum

There was no significant decrease in mean disease severity after 7 days. However significant reductions were obtained after 14 and 21 days and were statistically at par. There was no significant interaction of bacterial isolate with days after treatment for reduction of disease severity (Table 1).

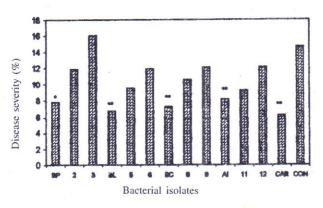
Table 1: Influence of bacterial isolates on severity and control of Myrothecium leaf spot in mulberry

Bacterial isolates	Di	Mean		
	7 DAS	14 DAS	21DAS	
Arthrobacter ilicis	17.28	13.99	13.17	14.81
	(-9.85)	(11.95)	(31.61)	(12.67)
Bacillus vereus	12.37	10.26	10.38	11.00**
	(21.36)	(35.43)	(46.10)	(35.00)
Bacillus lentimorbus	13.76	8.53	8.96	10.41**
	(12.52)	(46.31)	(53.47)	(38.07)
Bacillus pumilus	16.34	12.04	9.90	12.76*
	(-3.87)	(24.22)	(48.59)	(24.09)
Carbendazim (0.1%)	12.98	10.47	6.41	9.95**
	(17.48)	(34.10)	(66.71)	(40.80)
Check	15.73	15.89	19.26	16.96
	(0.00)	(0.00)	(0.00)	(0.00)
Mean	14.74	11.86**	11.35**	

^{*} Significant P, 0.05, ** P < 0.01, DAS = days after spray, Figures in parenthesis are percent disease control.

Significant reduction in mean disease severity was

obtained in plants treated with *B. cereus*, *B. lentimorbus* and *B. pumilus* as well as carbendazim lowering mean severity from 16.96 % to 11.89 %, 10.41 %, 12.76 % and 9.95 % respectively. The reduction in disease severity by *B. lentimorbus* and *B. cercus* was statistically as par with that of carbendazim. Compared to check, highest mean disease controlling efficacy showed by carbendazim (40.80 %). Except *A. ilicis*, all *Bacillus* isolates controlled the disease significantly with highest in *B. lentimorbus* (38.07 %) followed by *B. cereus* (34.56 %) and *B. pumilus* (24.09).



Mean of three replications, BP—B. pumilus, BL—B. lentimorbus, BC—B. cereus, AI—A. ilicis, CAR—carbendazim, CON—control. *Significant p < 0.05, ** significant p < 0.01

Fig. 2: Influence of bacterial isolates on severity of Myrothecium roridum

The rearing result showed no significant influence of bacterial treatment of the economic characters of silkworm (Table 2).

The results reported here indicate that bacterial isolates protect mulberry plants from *Myrothecium* leaf spot. The disease severity reduced significantly by treatment with *Bacillus isolates*, which was comparable to carbendazim. Several *Bacillus* strains are reported antagonistic to plant pathogens (Baker and Cooke, 1982; Ferreira *et al.*, 1991; Fravel and Spurr, (1977). The disease controlling efficiency of the bacterial isolates in the present study may be due to any one of the mechanisms such as the microbial parasitism, elicitation of phytoalexin, induction of systemic resistance, antibiotics produced by the bacterial isolates or a combined effect of these factors. Several workers (Thomashow and Waller, 1988; Kloepper, 1991;

Zhou et al., 1994) proved production of bacterial metabolites such as siderophores, hydrogen cynide, antibiotics, or extracellular enzymes that account for antagonism against the pathogens as mechanism of biocontrol. About 66 different antibiotics are reported to produce different *Bacillus* spp. (Katz and Demin. 1977). Significant reduction of disease was obtained in 14 and 21 days after treatment may be due to the increase in population of the bacterial isolates on the phyllosphere.

Table 2: Effect of feeding bacteria treated mulberry leaves to silkworm

Bacterial isolates	SCW(g)	SSW(g)	SR(%)	Wt. of 10 mature larvae (g)	ERR No.	Filament Length (mt.)
A. ilicis	0.9130	0.106	11.59	17.16	9800	369
B. cereus	0.9010	0.121	13.47	16.92	9400	360
B. lentimorbus	0.9047	0.1123	12.41	16.73	9667	367
B. pumilus	0.9540	0.1187	12.43	17.63	9670	362
Carbendazim	0.9517	0.117	12.33	17.03	9400	356
Check	0.8223	0.098	11.95	15.76	9733	343
CD (P < 0.05)	NS	NS	NS	NS	NS	NS

SCE: Single cocoon weight, SSW: Single shell weight, SR: Silk recovery (%), ERR: Effective rearing rate, NS: Not significant.

The studies are further required on the population dynamics of these isolates on the phyllosphere and the mechanism actually involved is whether induced resistance, micro-parasitism or a combined effect.

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REFERENCES

- Baker, K. F. and Cook, R. J. (1982). Biological control of plant pathogens. The American Phytopathological Society. St. Paul, MN. pp. 433.
- Govindaiah, Gunasekhar, V. and Sharma, D. D. (1988). Control of *Myrothecium roridum* causing leaf spot of mulberry. *CSR & TI, News Letter.* 3: 5-6.
- Elad, Y. and Steinberg, D. (1994). Effect of compost water extracts on grey mould (*Botrytis cinerea*). Crop. Prot., 13: 109-114.
- Ferreira, J. H. S.; Matthee, F. N. and Thomas, A. C. (1991).

 Biological control of *Eutipa lata* on grapevine by an antagonistic strain of *Bacillus subtilis*. *Phytopathology* 1: 283-287.

- Fravel, D. R. and Spurr, H. W., Jr. (1977). Biocontrol of tobacco brown spot disease by *Bacillus cereus subsp.* mycoides in a controlled environment. *Phytopathology*. 67: 930-932.
- Jolly, M. S. (1987). Appropriate sericulture techniques. International Centre for Training and Research in Tropical Sericulture, Mysore, India. pp. 175.
- Jongebloed, P. HJ.; Kessel, G. J. T.; Molhoek, W. ML.; van der Plas and Fokkeman, N. J. (1993). Biological control of *Phytophthora infestans* with compost extracts and selected bacterial antagonists. In: *Proceedings on Workshop on Biological control of Foliar and Post Harvest Diseases*, N. J. Fokkema, J. Kohl and Y. Elad (eds.), OBC/WPRS, Montfavet, France, pp. 16-22.
- Katz, E and Demin, A. C. (1977). The peptide antibiotic of Bacillus: Chemistry, biogenesis and possible functions. Bacteriol. Rev. 41: 449-474.
- Kloepper, J. W. (1991). Plant growth-promoting rhizobacteria as biological control agents of soilborne disease. In: The biological control of plant diseases. J. Bay-Peterson, (ed.). Food and Fertilizer Technology Centre, Taiwan. pp. 142-152.
- Philip, T.; Govindaiah, Bajpai, A. K. and Data, R. K. (1990). Chemical control of mulberry diseases in India-a review. *Indian J. Seric.* 33: 1-5.
- Pratheesh Kumar, P. M. (2002). Studies on fungal leaf spot diseases of mulberry (*Morus* spp.) in West Bengal. *Ph. D Thesis*, School of Life Sciences, Visva Bharati, Santiniketan, India. pp 224.
- Pratheesh Kumar, P. M. and Chattopadhyay, S (2002). Survey and Surveillance on the incidence of mulberry diseases in West Bengal. Annual Report. Central Sericultural Research and Training Institute, Berhampore, West Bengal. pp. 97-98.
- Pratheesh Kumar, P. M., Pal, S. C. and Qadri, S. M. H. (2002).

 Relation between leaf spot (Myrothecium roridum) intensity and physio-biochemical changes in mulberry (Morus spp.) National Symposium on Diversity of Microbial resources and their potential applications, 4-6, March 2002. Siliguri, India pp. 21. (abstr.).
- Pratheesh Kumar; P. M. Qadri; S. M. H., Maji; M. D. Gangwar; S. K. and Saratchandra, B. (2000). Biological control of *Phyllactinia corylea* (Pers.) Karst, using composted plant extracts. *Indian J., Seric.* **39**: 81-83.
- Qadri, S. M. H.; Gangwar, S. K.; Pratheesh Kumar P. M. Elangovan, C.; Das, N. K.; Maji, M. D. and Saratchandra, B. (1999). Assessment of cocoon crop loss due to leaf spot disease of mulberry. *Ind. J. Seric*. 38: 35-39.
- Shree, M. P. and Nataraja, S. (1993). Post infectional biochemical and physiological changes in mulberry. *Curr. Sci.* **65**: 337-341.
- Subba Rao, G. (1989). *Handbook of sericulture Extension and Management*. Central Sericultural Research and Training Institute, Berhampore, India. pp. 61.
- Teotia, R. S. and Sen, S. K. (1994). Mulberry diseases in India and their control. *Sericologia*. 34: 1-18.

- Thomashow, L. S. and Waller, D. M. (1988). Role of Phenazine antibiotic from *Pseudomonas fluorescence* in biological control of *Gaeumannomyces graminis* var. *tritici. J. Bacteriol.* 170: 3499-3508.
- Trankner, A. (1992). Use of agricultural and muncipal wastes to develop suppressiveness to plant pathogens. In: Biological control of plant diseases: Progress and Challenges for the future. E. C. Tjmos. G. c. Papavizas and R. J. Cook (eds.), Springer-Werley, New York. pp. 35-42.
- Umesh Kumar, N. N. (1991). Physiological studies of the mulberry varieties infected by foliar pathogens. Ph. D. Thesis. Bangalore University, Bangalore, India.
- Weltzien, H. C. (1991). Biological control of foliar fungal diseases with compost extracts. In: *Microbial ecology* of leaves. J. H. Andrews and S. S. Hirano, (cds.), Springer-Verlag, New York, pp. 430-450.
- Zhou, T., and Paulitz, T. C. (1994). Induced resistance in the biological control of *Pythium aphenidermatum* by *Pseudomonas* spp. *J. Phytopathol.* **142**: 51-63.

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