J. Mycopathol. Res. 38(2): 129–131 (2000) Printed in India © The Indian Mycological Society, Department of Botany, Calcutta University, Calcutta 700 019, India

Extent of lysis of *Rhizoctonia solani* Kühn cell wall preparation by different hyperparasites

ANJANI KUMAR SRIVASTAVA AND R.K. SINGH "KAVI"

Department of Botany, Ranchi University, Ranchi 834008, Bihar

Purified cell wall of *Rhizoctonia solani* was treated with enzyme preparation of wellknown hyperparasites of *Rhizoctonia solani* viz *Trichoderma harzianum*, *T. viride*, *T. hamatum* and *Aspergillus flavus*. The extent of lysis of the cell wall of *Rhizoctonia solani* due to the enzymes of hyperparasites was studied with the help of nephelometer. *T. harzianum* caused maximum lysis of 86% by 25 h followed by *T. viride* (45%), *T. hamatum* (15%) and *A. flavus* (6%). The optimum temperature for enzymic lysis of *Rhizoctonia solani* cell wall was found to be 20°C. The results indicate that *T. harzianum* can be used as biological control agent of *Rhizoctonia solani* in Chotanagpur plateau.

Key words: Rhizoctonia solani, lysis, hyperparasites, biological control agent

INTRODUCTION

Black surf, incited by *Rhizoctonia solani* Kühn, is a major disease of potato in the tribal region of Chotanagpur. The chemical measures to control the disease is not only costly but also less effective (Elad et. al., 1983). Thus another approach which could be used to control the disease is the biocontrol agents particularly *Trichoderma* spp. (Papavizas, 1985). Mycoparasitism is considered to be an important mechanism of biological control and probably depends on the production of lytic enzymes (Chet, 1987). This article reports the comparative abilities of *Trichoderma* spp. and *Aspergillus flavus* to cause lysis of cell walls of *Rhizoctonia solani*.

MATERIALS AND METHODS

Culture

Rhizoctonia solani Kühn (No. 2755) isolated from potato tubers were obtained from IARI, New Delhi. Following hyperparasites of *Rhizoctonia solani* were obtained from IARI, New Delhi : 1) *Trichoderma viride* (Pers) Gray (No. K-133), 2) *Trichoderma harzianum* Rifai (No. 1894), 3) *Trichoderma hamatum* (Bon) Bain (No. 2084), 4) *Aspergillus flavus* Link-ex. Fries (No. 315)

Preparation of Rhizoctonia solani cell wall

Rhizoctonia solani growing on PDA slants was harvested after 20 days of growth in 5 ml of distilled sterile water from each slant. An aliquot of 25 ml of Rhizoctonia solani suspension was obtained from 5 slants. They were crushed and homogenesized in a homogenizer (A. HT Philadelphia, U.S.A). It was washed by repeated centrifugation (3000 rpm) with O.1m NaCI, O.M acetate buffer (pH=5.5) and distilled water until the cells were free from cytoplasmic materials. The wall was inactivated by heating at 100°C for 30 minutes in a boiling water bath. An aliquot 0.2, μM of Na₂NO₃ was then added to keep the preparation sterile.

Preparation of enzyme

Trichoderma viride, Trichoderma hamatum, Trichoderma harzianum, and Aspergillus flavus were grown for 7 days at 25°C in stationary cultures in 250 ml conical flasks containing 100 ml of mineral medium of following composition (g/l) at pH = 5.5 : Glucose (anhydrous), 10; Ammonium tartrate, 2; K₂HPO₄, 1; MgSO₄, 7H₂O, 0.5 ; Leaf extract, 1; and Trace solution, 1 ml.

The trace element solution contained the following (in mg/l): Na₂B₄O₇, 10H₂O, 100; ZnSO₄, 7H₂O,

70; FeSO₄, 7H₂O, 50; CuSO₄, 5H₂O, 10; MnSO₄, 4H₂O, 10; (NH₄)₆ Mo₇O₂₄, 4H₂O, 10.

The culture filtrate was separated from the mycelium by filtration through Whatman Filter No. 3. The filtrate was centrifuged at 3000 rpm for 30 min. The culture filtrate was then dialysed twice against two changes of distilled water for 48 h at 4°C. This resultant preparation was taken as that of enzyme for studies on lysis of cell wall of *Rhizoctonia solani*.

Measurement of Lysis

One mg of Rhizoctonia solani cell wall preparation was taken in 1 ml of 0.05 M borate-citratephosphate buffer (pH=5.5). This was incubated for different time (1 h, 5 h, 10 h, 15 h, 20 h and 25 h and temperature (5°C, 10°C, 15°C, 20°C, 25°C, 30°C and 35°C) with 1 ml enzyme preparation. Turbidity was measured in Nephelometric turbidity units (NTU) by Nephelometric method with the help of a Nephelometer (Systronic, India). The sample was shaken thoroughly and air bubbles were allowed to escape. Sample was diluted with one or more volumes of turbidity free water until turbidity level fell down within limits of 30 to 40 NTU. Turbidity in original sample was calculated from the turbidity of diluted sample and the dilution factor was known. The sample was, then, transferred to the turbidometer tube and direct reading was taken on the scale. The calcution was made as following.

Nephelometric Turbidity Units = $A \times (B+C)/C$

Where A = NTU found in diluted sample, B=volume of dilution water in ml, C=sample volume taken for dilution in ml.

The percentage loss in turbidity was taken as measure of lysis of *Rhizoctonia solani* cell wall by enzymes of different hyperparasites. The boiled enzyme (30 min) was taken as control for the experiment.

RESULTS AND DISCUSSION

The degree of lysis of the cell wall of *Rhizoctonia* solani by enzymes of different hyperparasites is shown in Table 1. The enzymes of *Trichoderma* harzianum rapidly digested the cell wall of *Rhizoctonia solani*. It lysed 86% of the cell-wall by 25 h. *T.viride*, another hyperparasite, lysed only

45% of the cell wall in the same period whereas lysis brought about by *T. hamatum* and *Aspergillus flavus* was insignificant.

Table 1 : Degree of lysis (%) of *Rhizoctonia* solani cell wall preparation harvested after 20 days of growth on PDA slant at 30°C

Hyperparasites	Time (h)					
	1	5	10	15	20	25
Trichoderma viride	10	17	25	40	42	45
Trichoderma hamatum	8	13	13	13	15	15
Trichoderma harzianum	16	29	40	63	75	86
Aspergillus flavus	3	3	5	5	6	6

Results are mean of 3 replicates.

Bartnicki-Garcia (1973) who studied the cell wall chemistry of R. solani reported that cell walls mostly consist of glucans and 6-8% chitin. On the basis of this Hadar et al. (1979) suggested the role of lytic enzymes in degradation of R. solani. Our studies directly confirms the lysis of R. solani cell wall by enzymes of T.harzianum and T.viride as the inactivated wall preparation of R. solani was lysed enzyme preparation with active of the hyperparasites. This mechanism hyperparasitization by Trichoderma is different from that reported by Sawant and Mukhopadhyay (1990) where its volatile and non-volatile toxic metabolites played major role in controlling the damping off disease of sugarbeet.

Hadar et al. (1979) used T. harzianum to control damping off of bean and tomato plant seedlings caused by Rhizoctonia solani and Pythium aphanidermatum respectively. The strain of T. harzianum used by them extracellularly expleted β-1,3 glucanase and chitinase (Chet and Henis, 1969). Elad et al. (1983), too, reported lysed sites and penetration holes at the area of contact of R. solani hyphae parasitized by Trichoderma spp. They also detected,, with scanning electron microscope and fluorescence microscope, high amount of expletion of β-1, 3 glucanase and chitinase in dual agar cultures where T. harzianum parasitized Sclerotium rolfsiji. Relatively low levels of β-1, 3 glucanase and chitinase were secreted with either fungus alone. The enzymatic activity diminished when antagonism was prevented by cycloheximide. Similar mechanism of T. harzianum action on Rhizoctonia solani cell wall is indicated in our studies.

The light and pH are not the major factors affecting hyperparasitism in nature and especially in soil (Shigo and Hillis, 1973). These factors nevertheless can influence host-parasite relation *invitro*. In view of this, the studies with light and pH were not conducted here. The experiments with temperature revealed that optimum temperature for degradation of inactivated cell wall of *R. solani* by enzymes of *T. harzianum* was 20°C (Table 2). This indicates that the black scurf disease can effectively be controlled in field by adjusting the time of sowing of potato, when soil temperature is around 20°C.

Table 2 : Degree of lysis of *Rhizoctonia solani* cell wall by enzyme preparation of *Trichoderma harzianum* at different temperatures

Incubation Temperature (°C)	Lysis (%) of <i>Rhizoctonia</i> solani cell wall measured after 1 h of incubation			
5	2			
10	9			
15	12			
20	25			
25	20			
30	16			
35	14			

Results are mean of 3 replicates.

The potatoes which are normally sown in September-October (temperature around 25-30°C) give visible symptoms of *Rhizoctonia solani* infection in late November to early December when temperature ranges from 16-20°C. The experiments with temperature indicate that potato sowing may be delayed by a month (late November to early December) when the temperature is around 20°C. This will reduce the infection of *R. solani* as by delayed sowing its superinfection by *T. harzianum* can be encouraged at the initial stage of the growth of potato.

The present study, thus, reveals that *T. harzianum* controls the infection of *R. solani* by effectively lysing its cell wall through its enzymes. The delayed sowing of potato, when the temperature is around 20°C, encourages the superinfection of *R. solani* by *T. harzianum*. This indicates that *T. harzianum* can be used as biological control agent to curb *R. solani* infection on potato in Chotanagpur plateau.

REFERENCES

- Bartnicki-Garcia, S. (1973). Fungal cell wall composition. In *Handbook of Microbiology*, Chemical Rubber Co., Cleaveland, OH, **2**: 201-214.
- Chet, I (1987). *Trichoderma* application mode of action and potential as biocontrol agent of soilborne plant pathogenic fungi, in *Innovative Approaches to Plant Discase Control* ed. by I. Chet. John Wiley & Sons. New York, pp. 137-160.
- Chet, I and Henis, Y. (1969). Effect of catechol and sisodium EDTA on melanin content of hyphal and sclerotial wall of *Sclerotium rolfsii* Sacc. and the role of melanin in the susceptibility of these walls to β-1, 3 glucanase and chitinase. *Soil Biol. Biochem.* 1: 131-138.
- Elad, Y, Chet, I., Boyle, P. and Henis Y. (1983). Parasitism of *Trichoderma* spp on *Rhizoctonia solani* and *Sclerotium rolfsii* scanning microscopy and fluorescence microscopy. *Phytopathology*, **73**: 85-88.
- Hadar, Y., Chet, I and Henis, Y. (1979). Biological control of Rhizoctonia solani damping off with wheat bran culture of Trichoderma harzianum. Phytopathology, 69: 64-68.
- Papavizas, G. C. (1985). *Trichoderma & Gliocladium*: Biology, ecology and potential for biocontrol. *Ann. Rev. Phytopathol.*, **23**: 23-54.
- Sawant, I.S. and Mukhopadhyay, A.N. (1990). Intregration of metalaxyl with *Trichoderma harzianum* for the control of Pythium damping off in sugarbeet. *Indian Phytopath.* **43**(4): 535-541.
- Shigo, A.L. and Hillis (1973). Heartwood, discoloured wood and microorganism in living trees. *Ann Rev. Phytopathol.* 11: 197-222.

(Accepted for publication July 26 2000)