

## Evaluation of mulberry phylloplane bacteria for biological control of powdery mildew of mulberry caused by *Phyllactinia corylea*

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- In order to develop biological control of powdery mildew caused by *Phyllactinia corylea* (Pers) Karst, five mulberry phylloplane bacteria (MPB) viz., *Bacillus* sp., *Bacillus megaterium*, *Micrococcus* sp., *Brevibacterium lines* and *Serratia marcescens* were isolated on nutrient agar medium. Five MPB were screened *in vitro* against *P. corylea* conidia by slide germination method. Analysis of variance revealed that *Bacillus* sp., *B. megaterium*, *Micrococcus* sp., and *B. linens* inhibited conidial germination significantly. *Phyllactinia* mildew control efficacy of five MPB was tested in potted mulberry plants cv. S<sub>1</sub> under natural condition. Twenty four hour old bacterial culture suspension (1x10<sup>8</sup> colony forming unit/ml) was sprayed 30 days after pruning. Four MPB viz. *Bacillus* spp., *B. megaterium*, *Micrococcus* sp., and *B. lines* reduced the disease severity significantly and was at par with 0.2% Thiovit (micronized wettable sulphur 80 WP). Feeding of silkworm (*Bombyx mori* L) with MPB sprayed leaves revealed no harmful effect on silkworm in respect to larval weight, cocoon production, silk content, filament length and egg laying capacity of the female moth. Overall results indicated that phylloplane bacterial strains have an appreciable potential of biological control *Phyllactinia* mildew and are also safe to silkworm.

**Key Words :** Biological control, mulberry, *Phyllactinia corylea*, phylloplane bacteria, powdery mildew

### INTRODUCTION

Mulberry (*Morus* sp.), the sole food plant of silkworm (*Bombyx mori* L) is cultivated in large scale in the Gangetic plain of West Bengal for silkworm rearing since medieval period. Powdery mildew caused by *Phyllactinia corylea* (Pers) Karst is one of the major fungal diseases of mulberry. The disease regularly appears during winter, affecting leaf yield and quality. Chemical fungicides like Carbendazim 50 WP and Wettable sulphur effectively control the disease. Use of fungicides, has limitation due to their residual toxicity to silkworm, high cost, kill non-targeted beneficial phyllospheric and rhizospheric microorganisms, and may lead to development of resistant strains of pathogen. The protective action of leaf surface organisms against foliar pathogens has been reported by Fokkema *et al.* (1975) ; Kalita *et al.* (1996); Maji *et al.* (2001,

2002) ; Saikia and Chowdhury (1993) ; Spurr (1992) ; and Verma *et al.* (1983). Keeping this in view, phylloplane bacteria strains of S<sub>1</sub> mulberry cultivars are isolated and their efficacy are evaluated *in vitro* and *in vivo* against *P. corylea* and phylloplane sprayed leaves are fed to silkworm larvae to test its residual toxicity.

### MATERIALS AND METHODS

#### *Isolation of mulberry phylloplane bacteria*

Mulberry cv. S<sub>1</sub> leaves were collected in sterile polythene bags from Central Sericultural Research and Training Institute, Berhampore farm. Five leaf discs of 10 mm dia were taken from each leaf by cutting out aseptically by a sterile cork borer. Fifty discs were transferred into 100 ml sterile dist. water in a flask and was shaken for 20 min. Aliquots of 1

ml, 0.5 ml and 0.1 ml suspensions were transferred into sterile petriplates, containing Nutrient Agar medium (45-50°C). Plates were incubated at 25°C in inverted position for seven days. Bacteria were isolated and purified by streak method. Mulberry phyloplane bacteria (MPB) were identified by comparing morphology, staining, cultural and biochemical characteristics on the guidelines of Bergey's Manual of Determinative Bacteriology.

#### *In vitro screening of antagonistic MPB against P. corylea*

MPB were screened against *P. corylea* by slide germination methods. An aliquot of 0.2 ml of  $1 \times 10^8$  bacterial suspension was smeared on clean grease free microscopic slide and allowed to dry for 20 minutes at room temperature. Powdery mildew infected leaves were collected from green house and gently shaken on the treated slide to release mature conidia on the slide and incubated for 24 hrs and per cent in moist chamber. Conidial germination was recorded under 100X magnification and per cent of conidial germination was calculated by observing the number of conidia germinated in twenty microscopic fields. The per cent of conidial inhibition was calculated by following (Vincent, 1947) formula.

$$I = \frac{C-T}{C} \times 100$$

Where, I = Inhibition percentage, C = Conidial germination in control ; T = conidial germination in treatment.

#### *Field evaluation of MPB for control of P. mildew*

Five MPB strains were tested *in vivo* against mildew during peak season. Pot experiment was conducted in completely readomised design during November 2001 on mulberry cv. S<sub>1</sub>. MPB strains (48 hr old) grown on nutrient glucose agar were suspended in sterile dist water and were adjusted to standard turbidity (0.1 OD at 600 nm) to contain approximately  $10^8$  cfu/ml. MPB suspension (30 ml/plant) was sprayed on mulberry foliage 30 days after pruning with hand sprayer (Baygon make) at 4-5 PM. Micronised wettable sulphur fungicide (Thiovit 80 WP) was sprayed at 0.2% concentration and control plants were sprayed with water. Each

set of treatment was triplicated. Per cent disease index (PDI) data were recorded 20 and 30 days after spray (DAS).

Disease incidence was recorded by randomly selecting five branches from each plant. In each plant the total numbers of healthy and diseased leaves were graded into 0-5 scale [0=healthy leaf, 1 = 1-5% leaf area infected, 2 = 6-10% leaf area infected, 3 = 11-25% leaf area infected, 4 = 26-50% leaf area infected, 5 = 51 and above per cent leaf area infected]. per cent of disease index (PDI) was calculated according to FAO (1967) formula.

$$PDI = \frac{\text{Sum of all numerical rating}}{\text{Total no. of leaves observed} \times \text{maximum grade (5)}} \times 100$$

#### *Bioassay of MPB treated leaf*

The effect of feeding of MPB treated leaves on silkworm rearing, cocoon production, silk quality and egg laying capacity was assessed through bioassay (silkworm rearing). Aqueous suspension of MPB ( $1 \times 10^8$  cfu/ml) was sprayed 30 days after pruning. Sprayed leaves were fed to silkworm (Nistari Marked) after 10 days of application from the date of hatching up to spinning. Each of the treatment was replicated three times. Fifty larvae were taken in each replication. Date of important rearing, reeling and grainage parameters were recorded.

## RESULTS AND DISCUSSION

#### *In vitro screening of MPB against P. corylea*

Analysis of variance revealed that *Bacillus* sp., *Bacillus megaterium*, *Micrococcus* sp., *Brevibacterium lines* and *Serratia marcescens* inhibited *P. corylea* conidial germination significantly (Table 1). Among the bacterial strains *B. lines* inhibited 74.40% conidial germination followed by *Bacillus* sp. (62.20%). Antagonistic activity of several *Bacillus* spp. against plant pathogenic fungi reported by Baker and Cook (1982) ; Broadbent *et al.*, (1971) ; Chang and Kommedahl, (1968) ; Fravel and Spurr, (1977) ; Gregory *et al.*, (1984) ; Katz and Demain (1977) reported that *Bacillus* sp. produce at least 66 different antibiotics. The antagonistic activity in the present study may be attributed to the production of some antifungal



## REFERENCES

- Baker, K. F. and Cook, R. J. (1982). *Biological Control of Plant Pathogens*. The American Phytopathological Society, St. Paul, MN.
- Brian, P. W. (1957). The ecological significance of antibiotic production. Pages 168-188. In *Microbiology and Ecology*, (Williams, R.E.O and Spenceer, C. C. Eds.). Cambridge University Press, Cambridge.
- Broadbent, P. K., Baker, F. and Waterworth, Y. (1971). Bacteria and actinomycetes antagonistic to fungal root pathogens in Australia. *Aust. J. Biol. Sci.* **24** : 925-944.
- Chang, J. and Kommedahl, T. (1968). Biological control of seedling blight of corn by coating kernels with antagonistic microorganisms. *Phytopathology*. **58** : 1395-1401.
- Cook, J. R. (1980). Management of the associated microbiota. Page 145-166. In : *Plant Disease—An advance Treatise*. Vol. I (Horsfall, J. W and Cowling, E. B. Eds.). Academic Press, New York.
- Deacon, J. W. (1991). Significance and ecology in the development of biocontrol agents against soil borne plant pathogen. *Biocontrol Sci. Technol.* **1** : 5-20.
- FAO (1967). *Crop Losses Due to Diseases and Pests*. FAO, Rome.
- Ferreira, J. H. S., Mathee, F. N. and Thomas, A. C. (1991). Biological control of *Eutypa lata* on grapevine by an antagonistic strain of *Bacillus subtilis*. *Phytopathology*. **81** : 283-287.
- Fokkema, N. I. Van de Laar, J. A. J., Nelis-Blomberg, A. L. and Schippers, B. (1975). The buffering capacity of the natural mycoflora of rye leaves in infection by *Cochliobolus sativus*, and its susceptibility to benomyl. *Neth. J. Plant. Pathol.* **81** : 176-186.
- Fravel, D. R. and Spurr, H. W. Jr. (1977). Biocontrol of tobacco brown spot disease by *Bacillus cereus* sub sp. *mycoides* in a controlled environment. *Phytopathology*. **67** : 930-932.
- Gregory, G. F., Schreiber, L. R. and Ichada, J. (1984). Microorganisms antagonistic to or producing antibiotic inhibitory to *Ceratocystis ulmi*. (Abstr). *Phytopathology*. **74** : 804-805.
- Hall, T. J., Schreiber, L. R. and Leben, C. (1986). Effect of xylem colonizing *Bacillus* spp. on wilt in maples. *Plant Disease*. **70** : 521-524.
- Kalita, P., Bora, L. C. and Bhagabati, K. N. (1996). Phylloplane microflora of citrus and their role in management of citrus canker. *Indian Phytopath.* **49** : 234-237.
- Katz, E and Demain, A. C. (1977). The peptide antibiotics of *Bacillus* : chemistry, biogenesis, and possible function. *Bacteriol. Rev.* **41** : 449-474.
- Kloepper, J. W. (1991). Plant growth promoting rhizobacteria as biological control agents of soil borne diseases. Pages 142-152 In : *The Biological Control of Plant Diseases* (Bay-Peterson, J. Eds.). Food and Fertilizer Technology Centre, Taiwan.
- Liu, L., Kloepper, J. W. and Tuzun, S. (1995). Induction of systemic resistance in cucumber against bacterial angular leaf spot by plant growth promoting rhizobacteria. *Phytopathology*. **85** : 843-847.
- Maji, M. D., Qadri, S. M. H. and Pal, S. C. (2001). Evaluation of mulberry phylloplane microorganisms for biological control of bacterial leaf spot of mulberry caused by *Xanthomonas campestris* pv. *mori*. *Ind. J. April. Res.* (In press).
- Maji, M. D., Pratheesh Kumar, P. M., Chattopadhyay, S. and Saratchandra, B. (2002). Evaluation of mulberry phylloplane bacteria for biological control of leaf spot of mulberry caused by *Myrothecium roridum*. *Ind. J. April. Res.* (In press).
- Rothrock, C. S. and Gottlieb, D. (1984). Role of antibiosis in antagonism of *Streptomyces hygroscopicus* var. *geldanus* to *Rhizoctonia solani*. *Can. J. Microbiol.* **30** : 1440-1447.
- Saikia, P. and Chowdhury, H. D. (1993). Phylloplane microflora for the control of bacterial leaf blight of rice caused by *Xanthomonas oryzae* pv. *oryzae*. *Indian Phytopath.* **46** : 218-223.
- Spurr, H. W. Jr. (1972). Biological control of tobacco brown spot. *Phytopathology*. **49** : 755-756.
- Verma, J. P. Singh, R. P. Chowdhury, H. D. and Sinha, P. P. (1983). Usefulness of phylloplane bacteria in the control of bacterial blight of cotton. *Indian Phytopath.* **36** : 574-577.
- Vicent, J. M. (1947). Distortion of fungal hyphae in presence of certain inhibitors. *Nature*. **95** : 596.
- Wie, G., Kloepper, J. W. and Tuzun, S. (1991). Induction of systemic resistance of cucumber to *Colletotrichum orbiculare* by select strains of plant growth promoting rhizobacteria. *Phytopathology*. **81** : 1508-1512.

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