# Ecofriendly approach to microbial control

## S. SEN GUPTA, S. N. GHOSH, S. B. GHOSH\* AND A. K. DAS

Mycology & Plant Pathology Research Laboratory, P. G. Department of Botany, Presidency College, Kolkata 700 073, W. Bengal, India. \* Department of Botany, S. C. College, Habra 743268, W. Bengal, India.

In the present investigation three lichens, two pteridophytes and seven angiospermic plants were screened for their antimicrobial activities against four bacterial and eight fungal strains. The various solvent extracts of all these plants were found to be antimicrobially active.

## INTRODUCTION

Varieties of diseases of both plants and animals are caused by several pathogenic bacteria and fungi. Insect pests of important cultivars, pets and farm animals together is a major cause of concern of our time. In most of these clinical or pathological cases, a huge varieties of synthetic bactericides, fungicides, antibiotics and pesticides are used to control the diseases or are used as chemotherapeutic agents. The majority of these antimicrobial compounds and pesticides are toxic, in one form or other, to different biological systems and as such are the sources of constant threats to our environment. The progress in biotechnology coupled with the global environmental awareness, have prompted us to search for safe, eco-friendly, non-toxic and effective antimicrobial agents from the safest possible sources as plants. Screening and reviewing of plant extracts for their antimicrobial activities have extensively been carried out by several workers since middle of the twentieth century, till date (Osborn, 1943; Dhar et al., 1968; Ray and Majumder, 1976; Ghosh et al., 1980; Naqvi et al., 991; Cowan, 1999; Bhat and shukla, 2001; Sengupta et al., 2002; Banerjee and Mukherjee, 2003; Daswani and Bohra, 2003; Ghosh and Das, 2003).

In this investigation, an attempt has been made to test different groups of plants from West Bengal, India, for their antibacterial and antifungal activities.

## MATERIALS AND METHODS

The collected plant materials were thoroughly cleaned, dried (at 40°C), powdered and steeped separately in different pure solvents (Ethanol, Methanol and water only in the present sets of experiments) at different proportions (1g/10ml for Holarrhena antidysenterica and Ramalina sp.; and 1g/16ml for other plants). The aqueous extraction was prepared by maceration of the plant materials with additional water (1ml/g plant material), followed by filtration. The extracts concentrated (to 1/2 of its volume) by evaporating the solvents in a rotary evaporator. The extrcts from Usnea barbata, Peltigera sp., & Ramalina sp. (the lichens), Spheromeris chinensis, & Christiella sp. pteridophytes), and Holarrhena antidysenterica, Coriandrum sativum, Mentha piperita, Murraya exotica, Centella asiatica, Vitex negundo & Tephrosia candida (the angiosperms), were tested for their antimicrobial activities against Bacillus cereus, Bacillus subtilis, Staphylococcus aureus and Escherichia coli (the bacteria), and Aspergillus niger, Acremonium kiliense, Alternaria brassicicola, Alternaria triticina, Curvularia lunata, Fusarium udum, Macrophomina phaseolina and Rhizoctonia solani (the fungi).

Antimicrobial activities of these extracts were assayed by disc diffusion technique. The bacterial

On microbial control:

inocula, in the form of cell suspensions (106/ml), were prepared from 20-22 hr old nutrient broth cultures. The fungal inocula, as spore or mycelial suspensions (105-6/ml), were prepared by scrapping the surfaces of 90-95 hr old cultures on PDA and subsequently by suspending the scrappings in sterile water. In all the experimental sets, the plating medium for bacteria was Nutrient Agar (NA) and for fungi it was Potato Dextrose Agar (PDA), with 1.8% (w/v) agar in both, to which the bacterial or fungal inocula (as the case might have been) were incorported (1ml suspension/plate of 20 ml medium). Sterilized discs (diameter 6mm: made from Whatman No. 1 filter paper) soaked in the extract, were allowed to stand for 5 minutes and were then carefully placed in the freshly prepared agar plates seeded with the test organisms. The control plates, also seeded with the test organisms, received only the solvents in discs. The bacterial plates were incubated at 37°C for 24 hr and the fungal plates were incubated at 28°C for 96 hr. The diameter of the zone-of-inhibition thus developed around the paper discs, were measured and compared with the control sets.

#### RESULTS AND DISCUSSION

It was revealed from the zone-of-inhibition resulting from disc diffusion assay that, almost all the plant extracts tested so far have antibacterial activities against all the test bacteria. Moreover, a good number of plants, tested in this screening programme, showed antifungal activities as well against a number of test fungi (Table 1). The plants as *Christiella* sp. and *M. piperita*, although are fairly active against the test bacteria, did not have any antifungal property. The extracts from *C. sativum* and *M. exotica* showed feeble antifungal

Table 1: In vitro activities of the plant extracts against some bacteria and fungi

The plants (& parts used)	Extractio n Solvents	1.4	Diameter of inhibition zone against test organisms											
		Bacteria				Fungi								
		Вс	Bs	Sa	Ec	An	Ak	Ab	At	Cl	Fu	Mp	Rs	
U. barbata (thallus+ascocarp)	Ethanol	NP	X	33	25	1 <del>-</del>	NP	12	15	X	NP	NP	NP	
Peltigera sp. (thallus+ascocarp)	Ethanol	NP	X	9	14	3	NP	7	_	_	NP	NP	NP	
Ramalina sp. (thallus+ascocarp)	Ethanol	21	16	X	20	5 <del></del>	8	X	X	X	_	_	_	
S. chinensis Bory. (leaves)	Ethanol	NP	X		9	<u> </u>	NP	15		7	NP	NP	NP	
Christiella sp. (leaves)	Aqueous	16	X	8	8	1	_	X		Х	-	X	X	
H. antidysenterica (leaves) (L). wall	Ethanol	13	16	7	15		15	_	X	Х	_	_	7	
C. sativum L. (leaves)	Methanol	9	X	8	7	_	-	X	X	-	· —	X	7	
M. piperita L. (leaves)	Aqueous	8	7	X	7		X	_	. X	X	100000	X	_	
M. exotica L. (leaves)	Aqueous	8	8	7	7		-	X	_	X	2 <del></del>	X	7	
C. asiatica (L). Urban (leaves)	Ethanol	9	X	-	8	-		X	_	10	-	X	X	
V. negundo L. (leaves)	Ethanol	Х	16	17	_	X	_	14	X	Х	X	Х	_	
T. candida dc. (leaves)	Aqueous	_	Х	9	8		18	X	19	15	14	7	15	

Legends/key words: '—' =no inhibition; "x" = very poor inhibition; "NP" = Not Performed; BC = Bacillus cereus; Sa = Staphylococcus aureus; Ec = Escherichia coli; An = Aspergillus niger; Ak = Acremonium kiliense; Ab = Alternaria brassicicola; At = Alternaria triticina; Cl = Curvularia lunata; Fu = Fusarium udum; Mp = Macrophomina phaseolina; Rs = Rhizoctonia solani.

activities only against R. solani, whereas the Peltigera sp. was only active against the fungus A. brassicicola. However, none of the test plants could show any activity against the test fungus A. niger. The extracts of T. candida, H. antidysenterica and Ramalina sp. showed commendable activities against A. kiliense, while the fungus A. brassicicola was found to be very sensitive against A. kiliense, while the fungus A. brassicicola was found to be very sensitive against S. chinensis, V. negundo and U. barbata. The fungus A. triticina was well inhibited by T. candida and U. barbata. Similarly C. lunata, F. udum and R. solani were inhibited by T. candida, and C. lunata by C. asiatica. On the other hand, among bacteria, B. cereus showed very high sensitivity against Ramalina sp., H. antidysenterica, Christiella sp., C. sativum and C. asiatica; B. subtilis against Ramalina sp., H. antidysenterica and V. negundo.

Similarly, Staph. aureus was sensitive against U. barbata, V. negundo, T. candida and Peltigera sp. and E. coil was very sensitive towards U. barbata, Ramalina sp. H. antidysenterica, Peltigera sp. and S. chinensis. The extract of Usnea barbata, Peltigera sp. and Sphaeromeris chinensis have already been tested against the microorganisms, B. cereus, A. kiliense, F. udum, M. phaseolina and R. solani previously from this laboratory (Sengupta et al. 2002), so the experiment on this plant materials does not performed.

It is now evident from the results thus obtained that, the tested plant extracts have more antibacterial activities than antifungal, under the present experimental conditions, against the microorganisms tested so far. Nevertheless, the antimicrobial activities of any plant extract may be attributed to a single compound or there may be a synergistic effect of the active principles present in the extract. Of all, the extracts from U. barbata, Ramalina sp., H. antidysenterica and T. candida are found to be very promising because of their effectivencess against a greater number of microorganisms under this test condition.

Taking a cue from the present screening programme, further investigations, regarding the isolation, purification and characterization, are in progress.

#### ACKNOWLEDGEMENT

The authors are thankful to Professor T. B. Jha, Head, Department of Botany, Presidency College, for providing authors with his invaluable advices and infrastructural facilities throughout the tenure of this research work.

#### REFERENCES

- Banerjee, G. and Mukherje, A. (2003). Antibacterial activity of a common weed. *Portulaca oleracea* L., *Geobios.* 30(2-3): 143-144.
- Bhat, M. N. and Shukla, B. K. (2001). Evaluation of some leaf extracts against *Pythium aphanidermatum in vitro* and pot culture. *Ind. Phytopathol.* 54(3): 395-397.
- Gowan, M. M. (1999). Plant product as antimicrobial agents. Chem. Microbiol. Rev. 12(4): 564-582.
- Daswani, L. and Bohra, A. (2003). Antibacterial activity of various plant parts extrets of some spice plants against human pathogenic strains of *Staphylococcus aureus*. *Sci. & Cult.*, **69**(3-4): 155-156.
- Dhar, M. L. and Dhar, M. M. (1968). Screening of Indian plants for biological activity: Part I, *Ind. J. Exp. Biol.* 6: 232-247.
- Ghosh, S. B.; Gupta, S. Chandra, A. K. (1980). Antifungal activity in rhizomes of *Curcuma amada* Roxb., *Ind. J. Exp. Biol.* 18: 174-176.
- Ghosh, S. N. and Das, A. K. (2003). Evaluation of some plant extracts for biological control of common parasites of cash crops in Hoogly district of West Bengal, *J. Mycopathol. Res.* **41**(1): 83-85.
- Naqvi, S. A. H.; Khan, M. S. W. and Vohora, S. B. (1991). Antibacterial, antifungal and anthelmintic investigations of Indian medicinal plants, *Fitotetrapia*. **62**(3): 221-228.
- Osborn, E. M. (1943). On the occurrence of antibacterial substances in green plants, *Brit. Jour. Exp. Path.* **24**: 227-231.
- Ray, P. G. and Majumder, S. K. (1976). Antimicrobial activity of some Indian plants, Eco. Bot. 30: 317-320.
- Sengupta, S.; Das, A. K. and Ghosh, S. N. (2002). Biocidal activity of some plant extracts, *Jour. Hill Res.* 15(2): 99-101.

(Accepted for publication June 2, 2004)